The following samples were collected from Williamson Hall rooms Outside:

- 232
- 310
- 310D
- 312



Report for:

Mr. Chad Johnson Eastern Washington University EH&S, 002 Martin Hall Cheney, WA 99004

Regarding:

Project: WLM Hall EML ID: 2112486

Approved by:

Operations Manager Joshua Cox

Dates of Analysis: Spore trap analysis other particles-Supplement: 03-12-2019

Service SOPs: Spore trap analysis other particles-Supplement (EM-MY-S-1038) AIHA-LAP, LLC accredited service, Lab ID #102297

All samples were received in acceptable condition unless noted in the Report Comments portion in the body of the report. Due to the nature of the analyses performed, field blank correction of results is not applied. The results relate only to the samples as received. Sample air volume is supplied by the client.

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Aerotech Laboratories, Inc

Client: Eastern Washington University C/O: Mr. Chad Johnson Re: WLM Hall 1501 West Knudsen Drive, Phoenix, AZ 85027 (800) 651-4802 Fax (623) 780-7695 www.emlab.com

Date of Sampling: 03-06-2019 Date of Receipt: 03-08-2019 Date of Report: 03-12-2019

OTHER BIOLOGICAL PARTICLES REPORT: NON-VIABLE METHODOLOGY

Location:		35485: itside		35482: 310 D		35493: n 310		35661: n 312		35528: n 232
Comments (see below)		one		Jone Jone		lone		lone		lone
Lab ID-Version‡:	9993	5283-1	9993	5284-1	999	5285-1	9995	5286-1	9995287-1	
	raw ct.	particles/m3	raw ct.	particles/m3	raw ct.	particles/m3	raw ct.	particles/m3	raw ct.	particles/m3
POLLEN										
Grass (Poaceae)										
Mulberry (Morus)										
Oak (Quercus)										
Other					13	170				
Pine (Pinaceae)										
Ragweed (Ambrosieae)										
Sycamore (Platanus)										
OTHER PLANT										
Algae										
Diatoms										
Fern, moss, etc. spores										
Other (wood, trichomes, etc.)					21	280				
OTHER PARTICLES:										
ANIMAL										
Epithelial (skin) cells	3	40	53	2,800	166	44,000	33	1,800	79	1,100
Feather barbs					3	40				
Hair										
Insect parts					12	160				
Mites										
FUNGI										
Hyphal fragments	1	13			15	200			1	13
NON-BIOLOGICAL										
Cellulose fibers	3	40	32	430	81	11,000	21	280	8	110
Glass fiber					42	2,200				
Starch particles					15	200	7	93		
Synthetic fibers					15	200				
Background debris (1-4+)†	< 1+		2+		>4+		2+		1+	
Sample volume (liters)	75		75		75		75		75	
Comments.									v	

Comments:

The analytical sensitivity is the spores/m3 divided by the raw count. The limit of detection is the analytical sensitivity multiplied by the sample volume divided by 1000.

Carbonaceous particles include soot and other combustion products. In most instances a detailed analysis of soot can be accomplished using scanning electron microscopy.

Note: Interpretation is left to the company and/or persons who conducted the field work.

† Background debris is an indication of the amounts of non-biological particulate matter present on the slide (dust in the air) and is graded from 1+ to 4+ with 4+ indicating the largest amounts. To evaluate dust levels it is important to account for differences in sample volume.

A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".
Aerotech Laboratories, Inc
EMLab ID: 2112486, Page 2 of 2



Report for:

Mr. Chad Johnson Eastern Washington University EH&S, 002 Martin Hall Cheney, WA 99004

Regarding:

Project: WLM Hall EML ID: 2112486

Approved by:

Operations Manager Joshua Cox

Dates of Analysis: Spore trap analysis: 03-12-2019

Service SOPs: Spore trap analysis (EM-MY-S-1038) AIHA-LAP, LLC accredited service, Lab ID #102297

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Client: Eastern Washington University C/O: Mr. Chad Johnson Re: WLM Hall

1501 West Knudsen Drive, Phoenix, AZ 85027 (800) 651-4802 Fax (623) 780-7695 www.emlab.com

Date of Sampling: 03-06-2019 Date of Receipt: 03-08-2019 Date of Report: 03-12-2019

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:		35485: itside		35482: 310 D		35493: n 310		35661: n 312		35528: n 232
Comments (see below)		one		one		one		A		one
Lab ID-Version‡:	999	5278-1	9995	5279-1	9995280-1		9995281-1		9995	5282-1
Analysis Date:		2/2019		2/2019		2/2019		2/2019		2/2019
	raw ct.		raw ct.	spores/m3	raw ct.	spores/m3	raw ct.			spores/m3
Alternaria					7	93				
Ascospores	1	13			9	480				
Basidiospores	3	40			16	850			1	13
Bipolaris/Drechslera group		_			3	40				
Chaetomium					2	27				
Cladosporium	1	13			48	2,600				
Fusarium						,				
Myrothecium										
Nigrospora										
Other brown	2	27	1	13	4	53				
Other colorless										
Penicillium/Aspergillus types†					6	320				
Pithomyces										
Rusts										
Smuts, Periconia, Myxomycetes					71	9,500				
Stachybotrys										
Stemphylium										
Torula										
Ulocladium										
Zygomycetes										
Background debris (1-4+)††	< 1+		2+		>4+		2+		1+	
Hyphal fragments/m3	13		< 13		200		< 13		13	
Pollen/m3	< 13		< 13		170		< 13		< 13	
Skin cells (1-4+)	< 1+		1+		4+		1+		< 1+	
Sample volume (liters)	75		75		75		75		75	
§ TOTAL SPORES/m3		93		13		14,000		< 13		13

Comments: A) No spores detected.

 $^{++}Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.$

The analytical sensitivity is the spores/m^3 divided by the raw count, expressed in spores/m^3. The limit of detection is the analytical sensitivity (in spores/m³) multiplied by the sample volume (in liters) divided by 1000 liters.

For more information regarding analytical sensitivity, please contact QA by calling the laboratory. ‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

[†] The spores of Aspergillus and Penicillium (and others such as Acremonium, Paecilomyces) are small and round with very few distinguishing characteristics. They cannot be differentiated by non-viable sampling methods. Also, some species with very small spores are easily missed, and may be undercounted.



Report for:

Mr. Chad Johnson Eastern Washington University EH&S, 002 Martin Hall Cheney, WA 99004

Regarding:

Project: WLM Hall EML ID: 2112486

Approved by:

Operations Manager Joshua Cox

Dates of Analysis: Spore trap analysis: 03-12-2019

Service SOPs: Spore trap analysis (EM-MY-S-1038) AIHA-LAP, LLC accredited service, Lab ID #102297

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Client: Eastern Washington University C/O: Mr. Chad Johnson Re: WLM Hall Date of Sampling: 03-06-2019 Date of Receipt: 03-08-2019 Date of Report: 03-12-2019

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:		27335485				27335482				27335493		
		Outside				Rm 310	D			Rm 310		
Comments (see below)		None				None			None			
Lab ID-Version [‡] :		9995278-	1			9995279-	1		9995280-1			
Analysis Date:	03/12/2019				03/12/2019				03/12/2019	9		
Sample volume (liters)		75				75				75		
Background debris (1-4+) ^{††}		< 1+				2+				> 4+		
	raw ct.	Count/m3	DL/m3*	%	raw ct.	Count/m3	DL/m3*	%	raw ct.	Count/m3	DL/m3*	%
Hyphal fragments	1	13	13	n/a					15	200	13	n/a
Pollen									13	170	13	n/a
§ TOTAL FUNGAL SPORES	7	93	n/a	100	1	13	n/a	100	166	14,000	n/a	100
Alternaria									7	93	13	1
Ascospores	1	13	13	14					9	480	53	3
Basidiospores	3	40	13	43					16	850	53	6
Bipolaris/Drechslera group									3	40	13	< 1
Chaetomium									2	27	13	< 1
Cladosporium	1	13	13	14					48	2,600	53	18
Other brown	2	27	13	29	1	13	13	100	4	53	13	< 1
Penicillium/Aspergillus types									6	320	53	2
Smuts, Periconia, Myxomycetes									71	9,500	130	68
Stachybotrys												
Torula												
Ulocladium												
Zygomycetes												
Commonte												

Comments:

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

The analytical sensitivity/limit of detection is the Count/m³ divided by the raw count, expressed in Count/m³.

*The detection limit/limit of detection (DL) per cubic meter (m3) has been rounded to two significant figures to reflect analytical precision.

^{††}Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x".

§ Total Fungal Spores has been rounded to two significant figures to reflect analytical precision.

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Client: Eastern Washington University C/O: Mr. Chad Johnson Re: WLM Hall

Date of Sampling: 03-06-2019 Date of Receipt: 03-08-2019 Date of Report: 03-12-2019

SPORE TRAP REPORT: NON-VIABLE METHODOLOGY

Location:		27335661: Rm 312			27335528: Rm 232					
Comments (see below)		А			None					
Lab ID-Version [‡] :		9995281-1			9995282-1					
Analysis Date:		03/12/2019			03/12/2019					
Sample volume (liters)		75			75					
Background debris (1-4+)††		2+				1+				
	raw ct.	Count/m3	DL/m3*	%	raw ct.	Count/m3	DL/m3*	%		
Hyphal fragments					1	13	13	n/a		
Pollen										
§ TOTAL FUNGAL SPORES		< 13	n/a	100	1	13	n/a	100		
Alternaria										
Ascospores										
Basidiospores					1	13	13	100		
Bipolaris/Drechslera group										
Chaetomium										
Cladosporium										
Other brown										
Penicillium/Aspergillus types										
Smuts, Periconia, Myxomycetes										
Stachybotrys										
Torula										
Ulocladium										
Zygomycetes										

Comments: A) No spores detected.

Spore types listed without a count or data entry were not detected during the course of the analysis for the respective sample, indicating a raw count of <1 spore.

The analytical sensitivity/limit of detection is the Count/m^3 divided by the raw count, expressed in Count/m^3.

*The detection limit/limit of detection (DL) per cubic meter (m3) has been rounded to two significant figures to reflect analytical precision.

†Background debris indicates the amount of non-biological particulate matter present on the trace (dust in the air) and the resulting visibility for the analyst. It is rated from 1+ (low) to 4+ (high). Counts from areas with 4+ background debris should be regarded as minimal counts and may be higher than reported. It is important to account for samples volumes when evaluating dust levels.

‡ A "Version" indicated by -"x" after the Lab ID# with a value greater than 1 indicates a sample with amended data. The revision number is reflected by the value of "x". § Total Fungal Spores has been rounded to two significant figures to reflect analytical precision.

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Date of Sampling: 03-06-2019 Date of Receipt: 03-08-2019 Date of Report: 03-12-2019

MoldRANGETM, Local Climate; Extended Outdoor Comparison

Outdoor Location: 27335485, Outside

Fungi Identified	Outdoor		Туріса	l Outd	oor Da	ata for	:	Typical Outdoor Data for:					
	data			arch in W ab Local					The entire year in Washington [†] EMLab Local Climate code ¹				
		A Annu		A Elev.	, B Rain		o. Range	A Annu		, A Elev	., B Rain		p. Range
					=79)					•	=905)		
Project zip code 99004	spores/m3	very low	low	med	high	very high	freq %	very low	low	med	high	very high	freq %
Generally able to grow indoors*													
Alternaria	-	-	-	-	-	-	10	13	13	27	53	93	28
Bipolaris/Drechslera group	-	-	-	-	-	-	< 1	7	7	13	27	50	2
Chaetomium	-	-	-	-	-	-	6	7	13	13	23	40	6
Cladosporium	13	53	53	160	640	1,900	71	53	110	400	1,300	2,500	83
Curvularia	-	-	-	-	-	-	< 1	8	13	13	19	39	2
Nigrospora	-	-	-	-	-	-	< 1	-	-	-	-	-	2
Other brown	27	-	-	-	-	-	22	7	13	24	53	53	30
Penicillium/Aspergillus types	-	53	53	160	320	380	78	53	53	160	480	840	84
Stachybotrys	-	-	-	-	-	-	< 1	-	-	-	-	-	2
Torula	-	-	-	-	-	-	3	13	13	13	53	66	4
Seldom found growing indoors**													
Ascospores	13	53	53	210	640	1,000	53	53	89	270	1,000	1,700	78
Basidiospores	40	53	53	160	600	910	78	53	130	530	1,900	4,100	91
Rusts	-	-	-	-	-	-	< 1	13	13	26	53	100	18
Smuts, Periconia, Myxomycetes	-	13	21	27	53	99	37	13	27	160	1,000	2,000	67
§ TOTAL SPORES/m3	93												

¹EMLab Local Climate codes are a climate classification scheme for statewide geographic areas. The MoldRANGETM Local Climate report uses the sampling location zip code to identify the EMLab Local Climate code in that area. Using information available from the NOAA weather database, the EMLab Local Climate code sharpens the precision of the MoldRANGETM reporting system, providing more reliable estimates of the range and average concentrations of the different airborne fungal spore types for each region. Additional information on the EMLab Local Climate code system can be found on the last page of this report.

[†]The Typical Outdoor Data represents the typical outdoor spore levels across the state for the time period and EMLab Local Climate code indicated. The last column represents the frequency of occurrence. The very low, low, med, high, and very high values represent the 10, 20, 50, 80, and 90 percentile values of the spore type when it is detected. For example, if the frequency of occurrence is 63% and the low value is 53, it would mean that the given spore type is detected 63% of the time and, when detected, 20% of the time it is present in levels above the detection limit and below 53 spores/m3. These values are updated periodically and if not enough data is available to make a statistically meaningful assessment, it is indicated with a dash.

‡ n is the sample size used to calculate the MoldRANGETM Local Climate data summarized in the table.

* The spores in this category are generally capable of growing on wet building materials in addition to growing outdoors. Building related growth is dependent upon the fungal type, moisture level, type of material, and other factors. *Cladosporium* is one of the predominant spore types worldwide and is frequently present in high numbers. *Penicillium/Aspergillus* species colonize both outdoor and indoor wet surfaces rapidly and are very easily dispersed. Other genera are usually present in lesser numbers.

** These fungi are generally not found growing on wet building materials. For example, the rusts and smuts are obligate plant pathogens. However, in each group there are notable exceptions. For example, agents of wood decay are members of the basidiomycetes and high counts of a single morphological type of basidiospore on an inside sample should be considered significant.

§ Total Spores/m3 has been rounded to two significant figures to reflect analytical precision.

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Date of Sampling: 03-06-2019 Date of Receipt: 03-08-2019 Date of Report: 03-12-2019

Understanding EMLab Local Climate Codes

Outdoor airborne spore concentrations are strongly influenced by climate and weather patterns, often resulting in pronounced seasonal and diurnal cycles (Burge 1995). The seasonal climatic changes directly affect the growth cycle of plants, thereby influencing fungal growth, spore maturation, and release cycles. By evaluating outdoor spore concentrations across similar climatic zones rather than for the state as a whole, it is possible to provide a more representative estimate of typical outdoor spore levels and frequency of occurrence for different airborne fungal spore types in a given area.

The EMLab Local Climate code system is a novel and patent pending classification system that uses data from the NOAA - National Oceanic and Atmospheric Administration database to define unique climate regions by state. The following local climate variables, for each statewide zip code, are obtained from NOAA and assigned a letter code of A (above the statewide average for that variable) or B (below the statewide average for that variable):

- 1. Annual High Temperature
- 2. Elevation
- 3. Rainfall/Precipitation
- 4. Monthly Temperature Range

The result is a 4-character code assigned to each statewide zip code, referred to as the Local Climate Code. Below are some examples of decoded Local Climate Codes:

AAAA = Above avg. Annual High Temperature, Above avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range **AABB** = Above avg. Annual High Temperature, Above avg. Elevation, Below avg. Rainfall/Precipitation, Below avg. Monthly Temperature Range **BBAA** = Below avg. Annual High Temperature, Below avg. Elevation, Above avg. Rainfall/Precipitation, Above avg. Monthly Temperature Range

The actual outdoor air sample data from matching local climate codes in each state are then compiled in a manner relating typical spore concentrations and frequency of occurrence.

The NOAA local climate variables were selected by mapping data points from a subset of approximately 145,000 weather and geographic database entries to over 80,000 outdoor spore trap samples with known zip codes and assessing them using orthogonal array experimental design techniques. The results were then compared to the typical ranges of spore types found when grouping zip codes using the Koppen-Geiger climatic classification system; a commonly used climatic system that provides an objective numerical definition in terms of climatic elements such as temperature, rainfall, and other seasonal characteristics . The EMLab Local Climate codes showed improved granularity and refinement of the zip code groupings, implying a better representation of the expected range of spore types to be found within an individual zip code.

The values on this report were calculated by obtaining the four variables listed above from the over 585 million data points of weather and geographic information available in the NOAA database, and determining the frequencies and percentile values of spore types by utilizing over 180,000 EMLab P&K outdoor spore trap samples with known zip codes.

This report groups statewide zip codes in relation to these EMLab Local Climate codes and summarizes MoldRANGETM data by month and year within each EMLab Local Climate code.

References:

Burge, Harriet, A. Bioaerosols: Boca Raton: Lewis Publishers, pp. 163-171, 1995.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor data" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. In addition, EMLab P&K may not have received and tested a representative number of samples for every region or time period. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the use or interpretation of the data contained in, or any actions taken or omitted in reliance upon, this report.

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Date of Sampling: 03-06-2019 Date of Receipt: 03-08-2019 Date of Report: 03-12-2019

MoldSTATTM: Supplementary Statistical Spore Trap Report

Outdoor Summary: 27335485: Outside

Species detected		Outdoor sample spores/m3			Typical outdoor ranges	Freq.
	<100	1K	10K	>100K	(North America)	%
Ascospores				13	13 - 240 - 6,600	77
Basidiospores				40	20 - 480 - 24,000	91
Cladosporium				13	27 - 520 - 9,000	89
Other brown				27	7 - 22 - 160	27
Penicillium/Aspergillus types				< 13	13 - 190 - 2,700	66
Smuts, Periconia, Myxomycetes				< 13	7 - 53 - 1,100	66
Total				93		

The "Typical outdoor ranges" and "Freq. %" columns show the typical low, medium, and high spore counts per cubic meter and the frequency of occurrence for the given spore type. The low, medium, and high values represent the 2.5, 50, and 97.5 percentile values when the spore type is detected. For example, if the low value is 53 and the frequency of occurrence is 63%, it would mean that we typically detect the given spore type on 63 percent of all outdoor samples and, when detected, 2.5% of the time it is present in levels below 53 spores/m3.

Indoor Samples

Location: 27335482: Rm 310 D

% of outdoor total spores/m3	Friedman chi- square* (indoor variation)		ent ratio** c/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)	
Result: 13%	dF: 3 Result: 16.3000 Critical value: 7.8147 Inside Similar: No	Result: 0.4000		dF: 4 Result: 0.4500 Critical value: N/A Outside Similar: N/A	Score: 105 Result: Low	
Species 1	Detected			Spores/m3		
		<100	1K	10K	>100K	
	Other brown				13	
	Total				13	

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Date of Sampling: 03-06-2019 Date of Receipt: 03-08-2019 Date of Report: 03-12-2019

MoldSTATTM: Supplementary Statistical Spore Trap Report

Location: 27335493: Rm 310

% of outdoor total spores/m3	Friedman chi- square* (indoor variation)	Agreement ratio** (indoor/outdoor)		Spearman rank correlation*** (indoor/outdoor)		MoldSCORE**** (indoor/outdoor)		
Result: 15013%	dF: 3 Result: 16.3000 Critical value: 7.8147 Inside Similar: No	Result: 0.6154		dF: 9 Result: 0.3542 Critical value: 0.5833 Outside Similar: No		Score: 300 Result: High		
Species 1	Detected			Spore	s/m3			
_		<100	1K	_	10K	>10	0K	
	Alternaria						93	
	Ascospores						480	
	Basidiospores						850	
Bip	olaris/Drechslera group						40	
	Chaetomium						27	
	Cladosporium						2,600	
Other brown							53	
Penicillium/Aspergillus types							320	
Smuts, Periconia, Myxomycetes							9,500	
	Total						14,000	

Location: 27335661: Rm 312

% of outdoor total spores/m3	Friedman chi- square* (indoor variation)	Agreement ratio** (indoor/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)			
Result: < 1%	dF: 3 Result: 16.3000 Critical value: 7.8147 Inside Similar: No	Result: 0.0000	dF: N/A Result: N/A Critical value: N/A Outside Similar: N/A	Score: 100 Result: Low			
Species 2	Detected	Spores/m3					
		<100 1K	10K	>100K			
	None Detected			<13			

Location: 27335528: Rm 232

% of outdoor total spores/m3	Friedman chi- square* (indoor variation)		ment ratio** or/outdoor)	Spearman rank correlation*** (indoor/outdoor)	MoldSCORE**** (indoor/outdoor)		
Result: 13%	dF: 3 Result: 16.3000 Critical value: 7.8147 Inside Similar: No	Result: 0.4000		dF: 4 Result: 0.8500 Critical value: N/A Outside Similar: N/A	Score: 101 Result: Low		
Species	Detected	Spores/m3					
		<100	1K	10K	>100K		
	Basidiospores				13		
	Total				13		

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Date of Sampling: 03-06-2019 Date of Receipt: 03-08-2019 Date of Report: 03-12-2019

MoldSTATTM: Supplementary Statistical Spore Trap Report

* The Friedman chi-square statistic is a non-parametric test that examines variation in a set of data (in this case, all indoor spore counts). The null hypothesis (H0) being tested is that there is no meaningful difference in the data for all indoor locations. The alternative hypothesis (used if the test disproves the null hypothesis) is that there is a difference between the indoor locations. The null hypothesis is rejected when the result of the test is greater than the critical value. The critical value that is displayed is based on the degrees of freedom (dF) of the test and a significance level of 0.05.

** An agreement ratio is a simple method for assessing the similarity of two samples (in this case the indoor sample and the outdoor summary) based on the spore types present. A score of one indicates that the types detected in one location are the same as that in the other. A score of zero indicates that none of the types detected indoors are present outdoors. Typically, an agreement of 0.8 or higher is considered high.

*** The Spearman rank correlation is a non-parametric test that examines correlation between two sets of data (in this case the indoor location and the outdoor summary). The null hypothesis (H0) being tested is that the indoor and outdoor samples are unrelated. The alternative hypothesis (used if the test disproves the null hypothesis) is that the samples are similar. The null hypothesis is rejected when the result of the test is greater than the critical value. The critical value that is displayed is based on the degrees of freedom (dF) of the test and a significance level of 0.05.

**** MoldSCORETM is a specialized method for examining air sampling data. It is a score between 100 and 300, with 100 indicating a greater likelihood that the airborne indoor spores originated from the outside, and 300 indicating a greater likelihood that they originated from an inside source. The Result displayed is based on the numeric score given and will be either Low, Medium, or High, indicating a low, medium, or high likelihood that the spores detected originated from an indoor source. EMLab P&Kreserves the right to, and may at anytime, modify or change the MoldScore algorithm without notice.

Interpretation of the data contained in this report is left to the client or the persons who conducted the field work. This report is provided for informational and comparative purposes only and should not be relied upon for any other purpose. "Typical outdoor ranges" are based on the results of the analysis of samples delivered to and analyzed by EMLab P&K and assumptions regarding the origins of those samples. Sampling techniques, contaminants infecting samples, unrepresentative samples and other similar or dissimilar factors may affect these results. With the statistical analysis provided, as with all statistical comparisons and analyses, false-positive and false-negative results can and do occur. EMLab P&K hereby disclaims any liability for any and all direct, indirect, punitive, incidental, special or consequential damages arising out of the data contained in, or any actions taken or omitted in reliance upon, this report.